

SIMULTANEOUS SEPARATION OF ENANTIOMERS OF DIASTEREOMERS BY LIPASES

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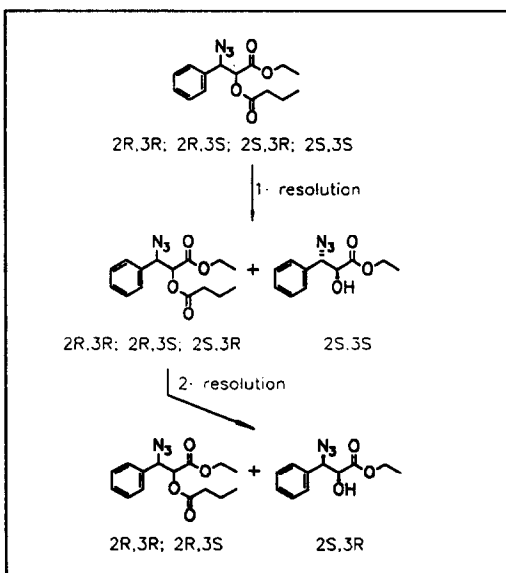
Abstract: Enantiomerically and diastereomerically pure 3-azido-2-hydroxy-3-phenyl propanoates are obtained from a mixture of racemic *threo*- and *erythro*-3-azido-2-butanoyloxy-3-phenyl propanoates by asymmetric hydrolysis with lipases.

Enzymatic resolution of diastereomerically pure educts is a well established procedure for the preparation of enantiomerically pure products.¹ In our continuing efforts to prepare rare amino acids in optical pure form by enzymatic methods, we chose 3-phenylserines and 3-phenylisoserines as model substrates for our approach to the class of hydroxy amino acids.² Recently we reported on the syntheses of enantiomerically pure 3-phenylisoserines by enzymatic hydrolyses of diastereomerically pure 3-azido-2-butanoyloxy-3-phenyl propanoates and subsequent hydrogenation of the products obtained.³

These syntheses involve tedious separations (distillations) of a mixture of *cis/trans* ethyl phenyl glycidates, which serve as starting materials for the preparation of 3-azido-2-hydroxy-3-phenyl propanoates, because mixtures of both *threo/erythro*-2-azido-3-hydroxy-3-phenyl propanoates and *threo/erythro*-3-azido-2-hydroxy-3-phenyl propanoates cannot be separated by column chromatography.

Here we want to describe the separation of (2*S*,3*S*)-3-azido-2-hydroxy-3-phenyl ethyl propanoate and (2*S*,3*R*)-3-azido-2-hydroxy-3-phenyl ethyl propanoate from a diastereomeric mixture of racemic *threo/erythro*-3-azido-2-butanoyloxy-3-phenyl propanoates⁴ by enzymatic resolution with lipases from *Pseudomonas* sp. and *Candida cylindracea*.

Only few literature examples are available on the diastereomeric separation with lipases. Sicsic et al.⁵ reported on a separation of diastereomeric norbornenyl esters by enzymatic resolution with pig liver esterase, but without enantiodifferentiation. Chenevert and Letourneau⁶ found that *erythro*-*N*-acetyl *p*-nitrophenylserinates can be resolved by enzymatic resolution with α -chymotrypsin, whereas *threo*-*N*-acetyl *p*-nitrophenylserinates were not hydrolyzed at all. In our case the *threo*-3-azido-2-butanoyloxy-3-phenylpropanoate worked in the enzymatic resolution with lipases as well, but about 20 times slower than the *erythro*-isomer. So it was possible to hydrolyze the (2*S*,3*S*)-isomer in the first resolution step together with only negligible amounts of the (2*S*,3*R*)-3-azido-2-hydroxy-3-phenyl ethyl propanoate. After chromatographic separation of the (2*S*,3*S*)-alcohol the remaining ester now containing the (2*R*,3*R*)-, the (2*R*,3*S*)- and the (2*S*,3*R*)-isomer was submitted to a further resolution. Thus the (2*S*,3*R*)-alcohol could be obtained after chromatographic purification. It was very important to determine the ratios of the isomers by e.g. ¹H-NMR prior to hydrolysis in order to know when the hydrolysis had to be stopped. Longer reaction times in the first resolution step decreased the diastereomeric excess (*de*) and enantiomeric



excess (*ee*) of the (2*S*,3*S*)-isomer, while increasing the *de* and the *ee* of the (2*S*,3*R*)-isomer, which is obtained in the second step (and *vice versa*). Results of the hydrolyses are shown in table 1 and 2.

experiment	erythro-alcohol							remaining ester				
	starting ratio ^b	time	conv. ^c	yield ^d	<i>de</i> ^b	<i>ee</i> ^e	config.	$[\alpha]_D^{20}$ ^f	yield ^d	ratio ^b	isomers	$[\alpha]_D^{20}$ ^f
No.	<i>threo/erythro</i>	h	%	%	%	%		%	<i>threo/erythro</i>			
1	80:20 ^g	3	10	9	93	59	2 <i>S</i> ,3 <i>S</i>	+50.3	86	87:13	2 <i>R</i> ,3 <i>R</i> ; 2 <i>S</i> ,3 <i>R</i> ;2 <i>R</i> ,3 <i>S</i>	-6.8
2	34:66	3	32	29	98	98	2 <i>S</i> ,3 <i>S</i>	+85.6	64	50:50	2 <i>R</i> ,3 <i>R</i> ; 2 <i>S</i> ,3 <i>R</i> ;2 <i>R</i> ,3 <i>S</i>	-36.0

Table 1: Results of the first resolution^a

experiment	<i>threo</i> -alcohol							remaining ester				
	starting ratio ^b	time	conv. ^c	yield ^d	<i>de</i> ^b	<i>ee</i> ^e	config.	$[\alpha]_D^{20}$ ^f	yield ^d	ratio ^b	isomers	$[\alpha]_D^{20}$ ^f
No.	<i>threo/erythro</i>	h	%	%	%	%		%	<i>threo/erythro</i>			
1	87:13	64	39	27	97	82 ^h	2 <i>S</i> ,3 <i>R</i>	-108.9	60	82:18	2 <i>R</i> ,3 <i>R</i> ; 2 <i>R</i> ,3 <i>S</i>	+20.7
2	50:50	65	20	17	90	94	2 <i>S</i> ,3 <i>R</i>	-124.5	75	44:56	2 <i>R</i> ,3 <i>R</i> ; 2 <i>R</i> ,3 <i>S</i>	-29.0

Table 2: Results of the second resolution^a

^a All reactions carried out in 100 ml 0.1M phosphate buffer at pH = 7.00, 1.00g substrate, 0.30 g enzyme (Lipase P from Amano, except were indicated), titration with 1N NaOH; ^b determined by ¹H-NMR (integration of the protons at C2 and C3); ^c measured by consumption of 1N NaOH; ^d isolated yield; ^e measured by ¹H-NMR and ¹⁹F-NMR of the respective MTPA-esters; ^f c = 2, CH₂Cl₂; ^g prepared in the same way as described in ref.4, but the ethyl phenyl glycidate was distilled twice; ^h *Candida cylindracea* (CC) at pH 7.00 was used instead of *Pseudomonas*, CC gives less enantiodifferentiation in that case.

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- (a)Hönig, H.; Seuffer-Wasserthal, P.; Weber, H., in: "Aminoacids: Chemistry, Biology and Medicine", Lubec G.; Rosenthal G.A. (Eds), ESCOM Science Publishers B.V., Leiden, 1990, in press; (b)Hönig, H.; Seuffer-Wasserthal, P.; Weber, H., *Tetrahedron*, 1990, 46, submitted.
- This mixture is prepared as follows: Darzens reaction of benzaldehyde and ethyl chloroacetate in DMF with *t*-BuOK⁷ results in a mixture of *cis/trans*-ethyl phenyl glycidates in ratios from *cis/trans* = 60:40 to 30:70 depending on the reaction time and the temperature applied. Epoxide opening with azide anion⁸ and subsequent acylation⁹ yields the mixture mentioned above, with the *threo/erythro*-ratios indicated in table 1 and 2. NMR-data and physical constants of all products were identical to those give in ref.3b.
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